

Cancers **2015**, *7*, 1271–1291; doi:10.3390/cancers7030836

OPEN ACCESS
cancers

ISSN 2072-6694

www.mdpi.com/journal/cancers

Review

Targeting Protein Kinase C Downstream of Growth Factor and Adhesion Signalling

Catriona M. Dowling ^{1,2,†,*} and Patrick A. Kiely ^{1,2,†,*}

¹ Department of Life Sciences, Materials and Surface Science Institute and Stokes Institute, University of Limerick, Limerick 78666, Ireland

² Health Research Institute (HRI), University of Limerick, Limerick 78666, Ireland

[†] These authors contributed equally to this work.

* Authors to whom correspondence should be addressed; E-Mails: Catriona.Dowling@ul.ie (C.M.D.); Patrick.Kiely@ul.ie (P.A.K.); Tel.: +353-61-202811 (C.M.D. & P.A.K.); Fax: +353-61-331490 (C.M.D. & P.A.K.).

Academic Editor: Robert H. Weiss

Received: 27 March 2015 / Accepted: 3 July 2015 / Published: 15 July 2015

Abstract: The signaling outputs of Receptor Tyrosine Kinases, G-protein coupled receptors and integrins converge to mediate key cell process such as cell adhesion, cell migration, cell invasion and cell proliferation. Once activated by their ligands, these cell surface proteins recruit and direct a diverse range of proteins to disseminate the appropriate response downstream of the specific environmental cues. One of the key groups of proteins required to regulate these activities is the family of serine/threonine intracellular kinases called Protein Kinase Cs. The activity and subcellular location of PKCs are mediated by a series of tightly regulated events and is dependent on several posttranslational modifications and the availability of second messengers. Protein Kinase Cs exhibit both pro- and anti-tumorigenic effects making them an interesting target for anti-cancer treatment.

Keywords: growth factor signaling; receptor tyrosine kinase; G-protein coupled receptors; integrins; Protein Kinase C; cell proliferation; cell adhesion.

1. Introduction

The composition and organization of the extra-cellular matrix (ECM) regulates cell behaviour and tissue morphogenesis. It does this by regulating integrin clustering and also by controlling the availability

of bioactives such as growth factors which control critical parameters such as cell migration, cell differentiation, cell polarity and cell proliferation [1,2]. The components of the ECM are regulated by hundreds of genes in the cells of the tissue and stroma [3]. Tumour cells respond to subtle changes in the composition of the ECM but can also influence the composition of the ECM by mediating dysregulation of particular sets of genes. This confers advantages to the tumour cells influencing changes in cytoskeletal dynamics, cortical tension, and microtubule turnover and enhances the transformed phenotype and promotes cancer progression [4]. How the cells detect and respond to changes in the ECM is mediated by a series of cell surface receptors that continuously sample the external environment. Signals from these cell surface receptors converge to recruit the cells adhesion and migratory machinery [5–7].

Receptor tyrosine kinases (RTKs) form a major part of the enzyme-linked family of receptors and include receptors for epidermal growth factor (EGF), fibroblast growth factor (FGF), platelet derived growth factor (PDGF), vascular endothelial growth factor (VEGF), hepatocyte growth factor (Met), glial cell line-derived neurotrophic factor (Ret), neurotrophins (Trks) and the insulin receptor family (IR) which includes the type I insulin-like growth factor receptor (IGF-1R) (reviewed in [5,8,9]). The activity of RTKs is under tight control by several modes of regulation such as transcriptional and post-transcriptional regulation and receptor internalisation. Once activated by their ligands, RTKs initiate diverse downstream signal transduction events to promote cell migration, but also proliferation, differentiation and the regulation of cell metabolism (reviewed in [5,8,10]).

Adhesion to the ECM is mediated by cell surface receptors such as integrins, members of the immunoglobulin superfamily and tyrosine kinase receptors [2]. Integrins are $\alpha\beta$ heterodimeric receptors that are composed of a large extracellular domain that binds ECM components and a short cytoplasmic tail that links to the actin cytoskeleton [11]. There have been 18 α subunits and 8 β subunits identified that associate with each other to generate 24 different receptors with distinct ligand specificities. For example, fibronectin which possesses both leucine-aspartic acid-valine (LDV) and arginine-glycine-aspartic acid (RGD) motifs can bind to many integrins (all five αV integrins, $\alpha 5\beta 1$, $\alpha 8\beta 1$, $\alpha 4\beta 1$, $\alpha 4\beta 7$ and $\alpha 9\beta 1$), laminin binds $\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 3\beta 1$, $\alpha 6\beta 1$, $\alpha 7\beta 1$, $\alpha 10\beta 1$, $\alpha 11\beta 1$ and $\alpha 6\beta 4$ and collagen binds $\alpha 1\beta 1$, $\alpha 2\beta 1$, $\alpha 10\beta 1$ and $\alpha 11\beta 1$ [12]. Integrins are the site of focal adhesion formation and these structures are required for polarised cell migration characterised by asymmetric adhesion dynamics with formation of adhesions at the leading edge and disassembly of adhesions at rear of the cell [13]. Like RTKs, integrins can independently propagate intracellular signals but cooperate with RTKs to initiate a cellular response through downstream signalling pathways to promote adhesion and migration.

There are many ways by which these receptor groups cooperate. Central to these events is the requirement for the integration of multiple signalling pathways. Clustering of integrins and phosphorylation of RTKs on specific tyrosine residues in the cytoplasmic domain creates binding sites for intracellular signalling molecules and facilitates the recruitment of adaptor proteins and specialized protein docking modules, including those with SH2 domains, plextrin homology (PH) domains, PDZ domains, and C2 domains (PKCs) [14]. Adaptor proteins in particular are strategically positioned at key steps in signalling pathways and function to disseminate and amplify signals accurately downstream of growth factor and adhesion receptors. Adaptor proteins generally do not possess enzymatic activity

but facilitate the creation of signalling complexes by bringing protein-binding partners together. The scaffolding of specific proteins in close proximity facilitates the reciprocal modulation of protein function and subsequent regulation of signalling events so that an appropriate response can be elicited.

An important function of activated cell surface receptors is the recruitment of intracellular kinases [5,15]. This recruitment leads to a cascade of downstream signalling and has a major influence on cell characteristics such as cell adhesion, proliferation, migration and invasion [16,17]. Focal adhesions are the converging point for growth factor and adhesion receptor signaling [18–20]. These are large dynamic macromolecular assemblies with signalling components and mechanical components and focal adhesions are assembled very precisely after the clustering of integrins on the cell surface. Integrin clustering is sufficient to promote the phosphorylation of focal adhesion kinase (FAK) on Tyr397. This facilitates the binding of the Src homology 2 (SH2) domains of Src family protein tyrosine kinases (Src-family PTKs) to promote the phosphorylation of FAK at secondary sites ensuring full activation of FAK. Once activated, FAK interacts directly with other non-receptor tyrosine kinases, cell surface receptors, cytoskeletal proteins and other adaptor proteins. We and others have characterised the interaction between Beta1 integrins and the IGF-IR and have shown that a WD repeat containing the scaffolding protein RACK1 mediates crosstalk between the IGF-IR and adhesion receptors by orchestrating the recruitment of a series of proteins to regulate focal adhesions [21–28].

A key subgroup of these intracellular kinases are Protein Kinase Cs (PKC), a family of serine/threonine kinases which play key roles in several signalling pathways [29–32]. This group of proteins are expressed in many different tissue types and hence have a diverse range of biological functions [33,34]. At focal adhesions, PKCs have been shown to interact with several structural, mechanical and regulatory proteins that are central to the establishment, maintenance and disassembly of focal adhesions (reviewed in [35]).

2. Protein Kinase C: Structure, Function and Activity

All members of the PKC family share common basic structures; a flexible hinge segment linking a cell membrane targeting N-terminal regulatory domain to a C-terminal catalytic domain [45,46]. The regulatory domain maintains the enzyme in an inactive conformation with two discrete membrane targeting modules, termed C1 and C2 [47]. The C1 domain binds diacylglycerol (DAG) in all but the atypical isozymes and the C2 domain binds anionic lipids with the conventional isozymes also binding phosphatidylserine (PS) and calcium [48–52]. The maturation of cPKCs into this inactive conformation is dependent on phosphorylation steps at three highly conserved sites termed: the activation loop, the turn motif, and the hydrophobic motif [53,54]. Phosphoinositide-dependent kinase-1 (PDK1) phosphorylates newly synthesised cPKC on a threonine residue at the activation loop which positions the active site for catalysis [47,55,56]. This phosphorylation triggers phosphorylation of the turn motif and consequently autophosphorylation of the hydrophobic motif, leading to the optimal stability of the enzyme [57,58]. The maturation of cPKC primes it for activation allowing changes in intracellular calcium levels to recruit the C2 domain to the membrane followed by the binding of DAG to the C1 domain and so the release of the active site [47] (Figure 1).

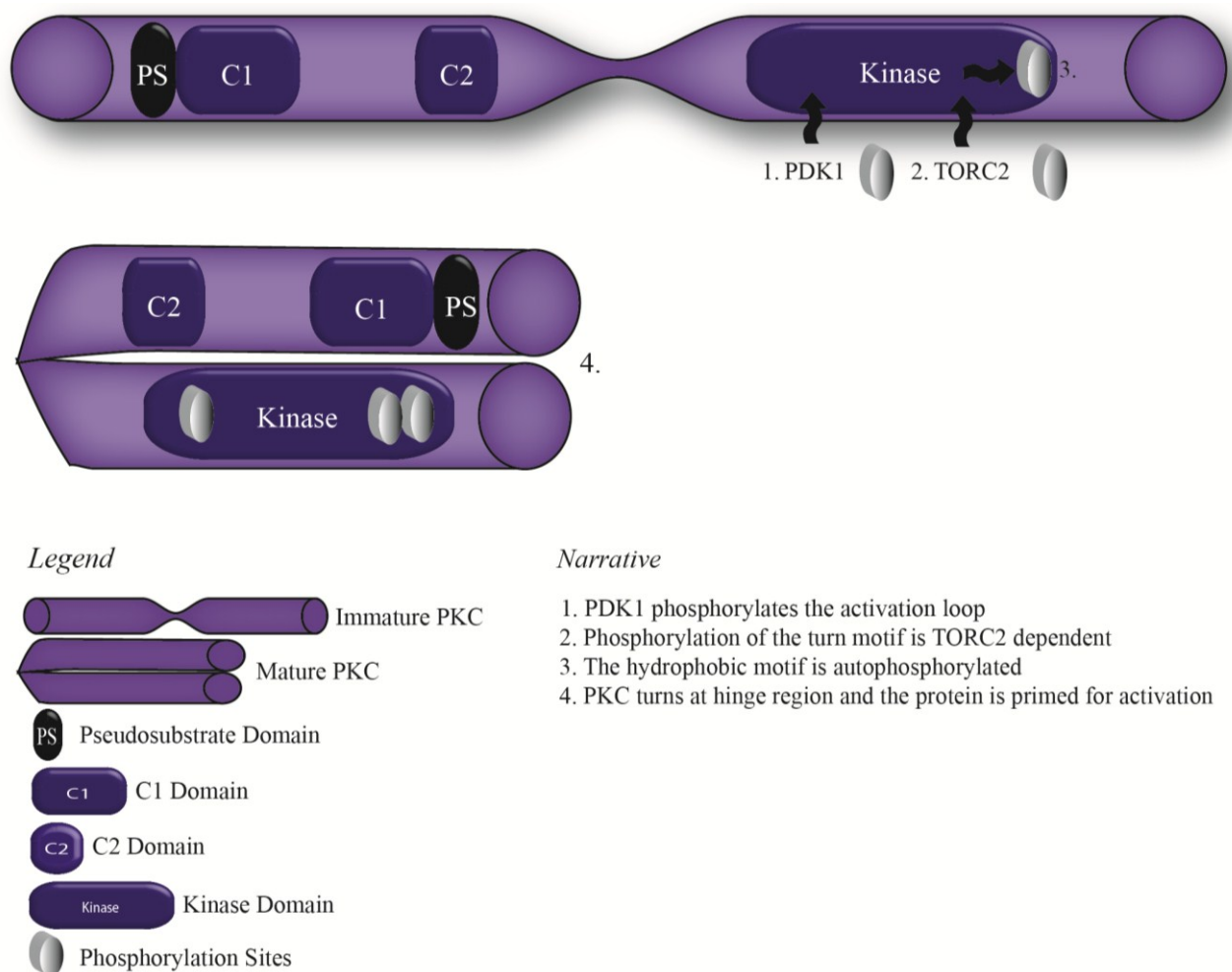


Figure 1. Priming of Protein Kinase C in to its inactive mature conformation.

3. Activation of PKCs: Receptor Tyrosine Kinases, G-protein Coupled Receptors and Integrins

The mechanisms underlying PKC activation have been well studied and are triggered by changes in intracellular cofactors of PKCs downstream of signals that stimulate RTKs, G-protein-coupled receptors (GPCR) and integrins [59].

There are 58 known human RTKs which fall into 20 subfamilies, many of which are believed to play a role in the activation of PKCs [8,9]. Growth factors were thought to activate various RTKs through dimerization but the intracellular signalling responses of RTKs were largely unknown. In 1984, EGF was found to stimulate the RAS oncoprotein [60] and shortly after, phospholipase c (PLC) was discovered as the first substrate that directly interacts with activated EGFR [61]. Dimerization of RTKs by growth factors causes the auto phosphorylation of their cytoplasmic tyrosine domains, creating a series of high affinity docking sites for signalling proteins such as phosphoinositide 3-kinase (PI3-kinase) and PLC [62,63]. PI3-Kinases are a large family of kinases consisting of three classes and multiple subunits [64]. They contain a regulatory and catalytic subunit; the p85 regulatory subunit either binds directly with activated RTKs through its SH2 domain or binding may be mediated through phosphoproteins such as the insulin receptor substrate [63,65]. PI3-kinase phosphorylates the 3 position hydroxyl group of the inositol ring of phosphatidylinositols (PtdIns) [66]. PLC isozymes

are divided into three classes, β , γ and δ , differing on their mechanisms of activation. PLC β ($\beta 1$ and $\beta 2$) is activated by G-proteins, little is known about the activation of δ isoymes ($\delta 1$ and $\delta 2$) and PLC γ ($\gamma 1$ and $\gamma 2$) is activated by RTKs [67]. PLC γ translocates from the cytosol to the membrane, where its N-terminal and C-terminal SH2 domain binds to the phosphorylated tyrosine site of the RTK [68]. Upon activation, PLC γ hydrolyse PtdIns generating two secondary messengers, namely, DAG and inositol phosphates (IPs). IPs stimulate the release of calcium from intracellular stores, activating cPKCs and translocation of the protein to the cell membrane at which the C2 domain can bind to PS, followed by the binding of DAG to the C1 domain (Figure 2) [69,70]. The binding of the C2 domain to PS in novel PKCs does not require calcium and thus their membrane recruitment and activation is calcium- independent [46]. This process of PKC activation has been well established for many growth factors such as EGF, PDGF and nerve growth factor (NGF) [59,70,71]. Following stimulation of PDGF receptors, PLC γ can bind to the receptor at tyrosine residue Y1021 and PI3-kinase can bind at tyrosine Y740 and Y751, resulting in the translocation of PKC ϵ [72,73]. PKC λ can also translocate from the nucleus to the cytosol when activated through growth factors PDGF and EGF but this translocation only involves PI3-kinase signalling [74]. EGF treatment of cells has also shown to translocate PKC α and PKC γ to the membrane with no effect on the translocation of PKC ζ [75,76]. Both EGF and NGF activation of PKC ϵ in neuronal cells provides a positive signal for neurite outgrowth [75,77].

The role of PKCs in regulating growth factor signalling has been well documented. For example, activation of PKCs via EGF results in the direct phosphorylation of the EGFR at Thr 654 leading to a decrease in ligand affinity and receptor activity [78–81]. It is believed that PKCs regulate whether the cellular response to EGF is pro-mitogenic or pro-motility [82–84]. PKC isozymes are also known to regulate the HGF receptor c-Met. PKC α controls the trafficking of c-Met to perinuclear compartments in a microtubule dependent manor [85] and PKC ϵ is required for the c-Met activation of ERK [86]. Activation of ERK downstream of PKC ϵ and their localization to focal adhesions is required for PKC ϵ induced adhesion and migration [87]. In contrast, it has also been shown that PKC δ and PKC ϵ can inhibit tyrosine phosphorylation of c-Met by phosphorylating c-Met at Thr 985 [59]. It is thought that these isozymes may be functioning to regulate c-Met receptor signalling as PKC δ and PKC ϵ mediated phosphorylation at Thr 985 is promoted by HGF itself and rapidly removed by protein phosphatase 2 (PP2A) [88].

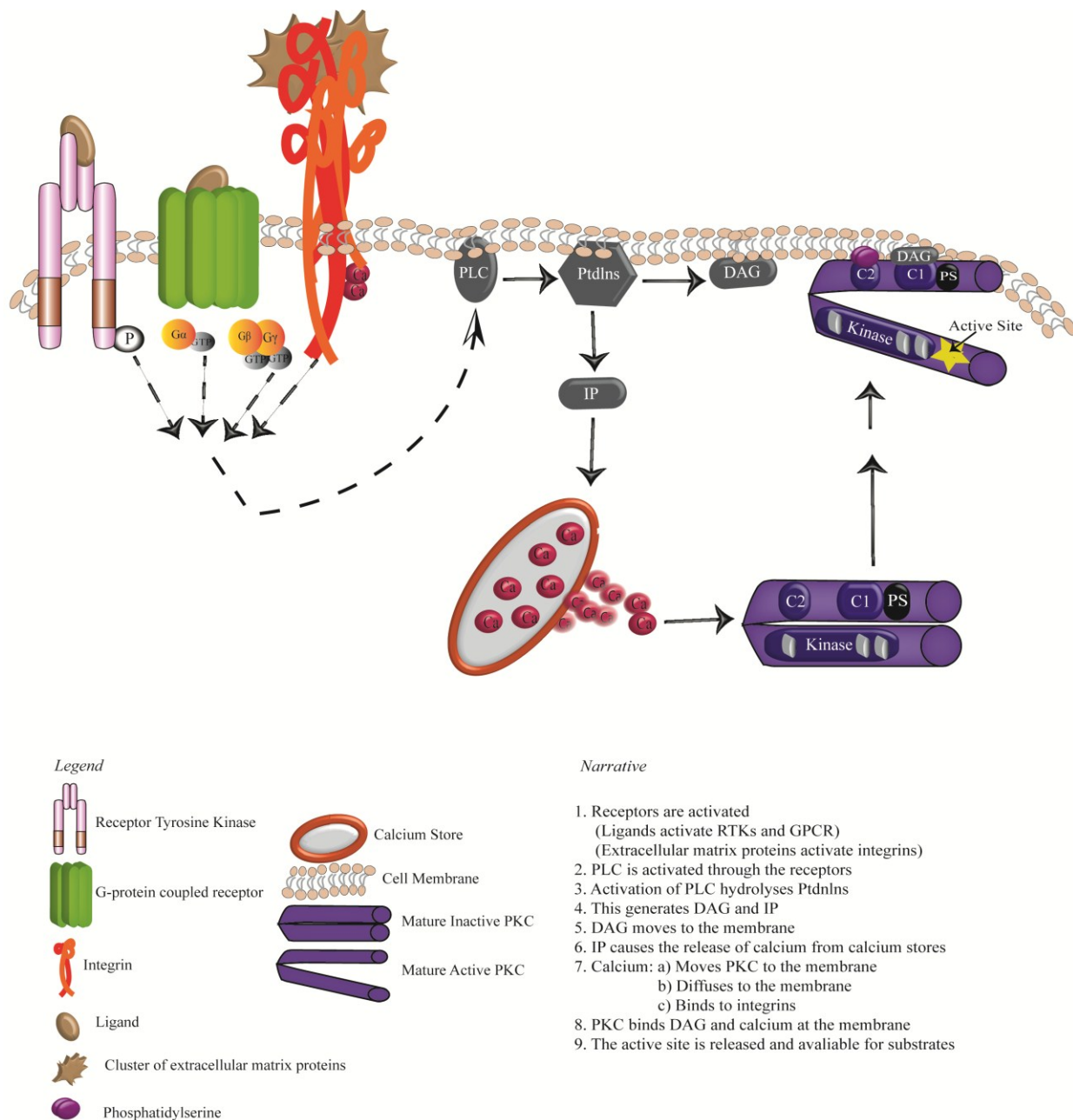


Figure 2. Activation of mature Protein Kinase C.

G-protein-coupled receptors have a characteristic core composed of seven transmembrane α helices weaving in and out of the membrane and with over 800 members they represent the largest family of cell-surface molecules involved in signal transmission [89,90]. Many different agonists stimulate GPCRs and upon stimulation of the extracellular side of the receptor introduce a conformational change allowing for the interaction of the heterotrimeric G-proteins with the intracellular sites on the receptor [91]. G-proteins contain three subunits; α , β and γ , when bound to the receptor guanosine diphosphate (GDP) association with the $G\alpha$ subunit is replaced with guanosine triphosphate (GTP), which in turn leads to the dissociation of $G\alpha$ from the $G\beta\gamma$ subunit and hence activation of the protein [92]. At present there are 17 known $G\alpha$, 5 β and 12 γ subunits with the $G\alpha$ subunits divided into four major families $G\alpha_s$, $G\alpha_i$, $G\alpha_q$ and $G\alpha_{12/13}$ [92,93]. The $G\alpha_q$ family play a major role in the activation of PKCs

through the stimulation of PLC- β hydrolysis of phosphatidylinositol biphosphate (PIP₂) producing inositol triphosphate (IP₃) and DAG, the two major secondary messengers required for PKC recruitment and activation (Figure 2) [89]. Traditionally it was thought that the G $\beta\gamma$ complex solely served to bind the G α subunit to prevent spontaneous signalling, however, in more recent years it has emerged as an activator of the PLC signalling pathway and hence, PKC activation (Figure 2) [94].

Protein Kinase C appears to be a key intermediate in integrin mediated signalling and while much of the research on integrins and PKCs has focused on the regulation of integrin activity by PKCs, recent studies also highlight that integrins play a central role in mediating activation of PKCs [95,96]. Extracellular matrix proteins cluster and activate integrins which in turn induce the PLC signalling cascade and the activation of PKCs, in a process termed outside-in signalling (Figure 2). This activation of PKCs results in a mobilization of calcium levels within the cell and consequently further activation of integrins, a phenomenon referred to as inside-out signalling [95,97–99]. Consequently, PKC activation has a central role to play in the establishment and maintenance of focal adhesions downstream of clustered integrins (reviewed in [35]). Inhibition of PKC activity in many cell types results in a reduction in cell spreading, a consequence of reduced focal adhesion formation in the cells [44,100].

4. Oncogenic Signalling Downstream of RTKS, GPCRs and Integrins

Research into PKC signalling intensified when it was discovered that PKC is a high-affinity intracellular receptor for phorbol-ester tumour promoters such as TPA [37,101,102]. This strongly suggested that PKC activation promoted tumorigenesis induced by carcinogens. PKC signalling and oncogenic signalling converge and contribute to the transformed phenotype [102–104], however it remains unclear as to whether tumour promotion is a result of changes in specific activity, or by changes in the expression of the protein [33,105–108]. Confusion also arises as immunohistochemical and biochemical studies indicate that altered expression of the PKC isozymes is variable and depends on the cancer cell type [32,42,59,109]. This could suggest that perhaps a change in both the activity and expression is associated with the transformed phenotype. In an attempt to delineate this confusion this review will discuss the most recent findings presented for each conventional isozyme.

When examining the expression levels of PKC α there are contradictory results emanating from different tissue types. Over expression of the isozyme is reported in tissue samples from prostate, endometrial, urinary bladder and hepatocellular cancers while down regulation has been observed in basal cell and colon cancer [110–116]. In breast cancer, up/down regulation of PKC α has been suggested to be dependent on the specific subtypes of the disease [117–119]. However, taken together, the majority of studies suggest PKC α plays a role in increasing the proliferation and invasive capacity of cancers and many PKC α inhibitors have shown to reverse the phenotype [120–122]. Studies propose PKC α facilitated invasion can occur in a number of different ways; through inhibition of protein complexes at cell junctions, inhibition and mobilisation of hemidesmosomes mediated through the β 4 integrin, and PKC α can introduce changes in β 1 integrin mediated cell matrix junctions [121,123,124]. More specifically in breast cancer, it has been proposed that alteration in the subcellular localization of PKC α results in a change in the desmosomal adhesive state of the cells potentially leading to a loss in cell–cell adhesion and a transition from a normal to a malignant phenotype [125].

Two splice variants have been described for PKC β and again the expression levels of both variants differ between tissue types. Loss of PKC β is observed in malignant melanocytes and melanoma cell lines [126]. The expression of PKC β 1 and β 2 in breast, gastric and colon cancer has been subject to much debate and there are many studies presenting arguments for both up and down regulation of the isozyme in the diseases [127–132]. Despite these conflicting studies, the role of PKC β in angiogenesis has been well documented. The isozyme plays a role in mediating VEGF signalling and its inhibition in this pathway results in decreased endothelial cell proliferation and reduction of neovascularization in malignant tumours [133,134].

PKC γ is predominantly expressed in neuronal tissues and there is very little evidence to suggest a role for PKC γ in tumorigenesis. In certain forms of B-cell lymphomas, PKC γ expression has proven to be a positive prognostic factor [135]. Mammary epithelial cells overexpressing PKC γ acquire a malignant phenotype *in vivo* [136], however, its role in breast cancer formation has not yet been established. More research into the role of PKC γ in tumour formation needs to be conducted in order to establish what role, if any, it plays in the development of tumours.

5. Consequences of Targeting PKC

There have been many efforts made in targeting PKCs for anti-cancer treatments [33,137]. A number of different approaches have been explored in an attempt to create selective modulators for the PKC isozymes including; ATP competitive small molecule inhibitors, phorbol esters and derivative activators and inhibitors which mimic the binding of diacylglycerol and peptides that disrupt protein-protein interactions between PKC and its corresponding RACK (Figure 3) [33,132,137,138].

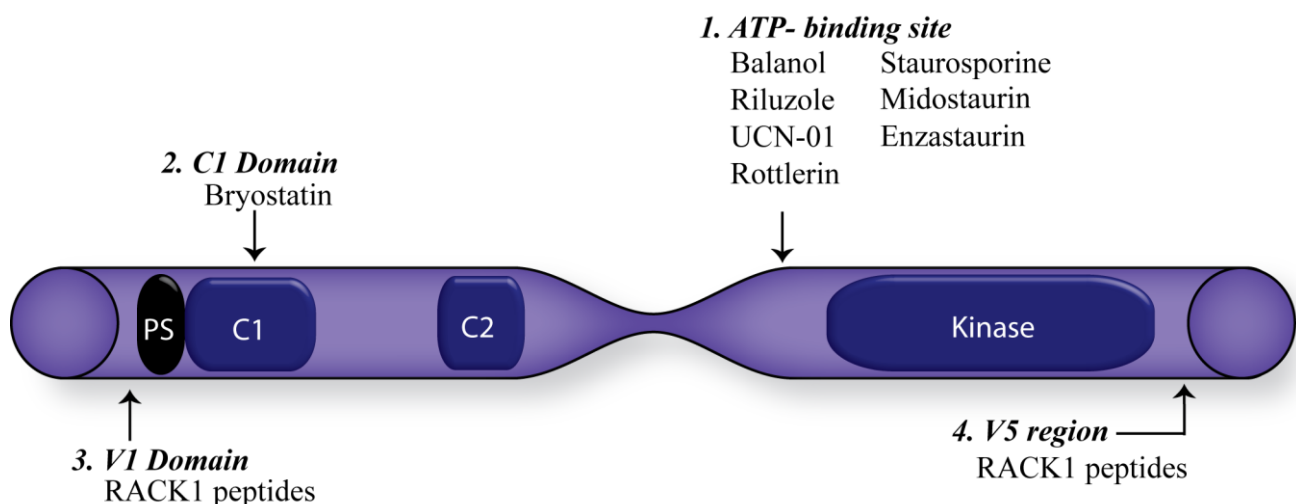


Figure 3. Therapeutic approaches targeting Protein Kinase C.

The first ATP-binding site inhibitor, staurosporine, was developed over 30 years ago. This compound binds to all PKC isozymes but also binds non-specifically to several other serine/threonine kinases [139]. Midostaurin (PKC412), a derivative of staurosporine, was subsequently developed in an attempt to design a more isozyme specific ATP-binding site inhibitor. The compound exhibits an increased specificity for the conventional and atypical PKCs but also inhibits other tyrosine kinase pathways [140]. In preclinical trials reports suggest that PKC412 may act as a radio-sensitizer for human xenografts

through the blockade of the P13K/Akt pathways [141]. Enzastaurin, emerged as another ATP-binding compound, thought to be specific to PKC β . While demonstrating a 20-fold more powerful inhibitor of PKC β , it is now believed that it can also inhibit other PKC isozymes [142]. Enzastaurin prevents angiogenesis through alteration of the VEGFR signalling cascade [143] and is currently in clinical trials for brain malignancies [144].

The C1 domain of the regulatory region of PKC's has also been targeted. A number of compounds have been developed over the years but the most famous of these compounds, bryostatin, a naturally occurring macrolactone, mimics the binding of DAG and has pan PKC activity [145]. Preclinical trials looked promising with bryostatin showing an effect on tumours including melanoma, leukemia, lymphoma and lung cancer [146]. Despite this, the drug has proven to be disappointing in clinical trials, as it showed very little efficacy in reducing tumour growth, even when combining the drug with other cytotoxic drugs and has since been suspended [37].

The translocation of PKCs to their subcellular locations is an imperative part of PKC activity, leading to the development of modulators that target the interaction sites of these shuttle proteins. The C2 domain is the main focus for designing inhibitors against protein-protein interactions; this is owed to the founding work of the Mochley-Rosen lab, who demonstrated that unique sequences within the C2 domain are a “hot-spot” for several protein-protein interactions [147]. However, other protein-protein interactions have been delineated in regions between the C1, C2, C3, and C4 regions, the V2, V3 and V5 regions as well as between the C1a and C1b subdomains [33]. These protein-protein interactions are the subject of a new generation of PKC targeting. For example, a peptide derived from the V5 region, which inhibits the interaction of PKC β II and RACK1 has shown to prevent cardiac dysfunction and death in rat models of post heart failure and inhibit neoangiogenesis in a xenograft mouse model of prostate cancer [148,149].

6. Conclusions

The role RTKs, GPCRs and integrins play in activating PKCs is well characterised and is a key feature of cell proliferation and oncogenic signalling. Traditionally, research into the role of different PKC isozymes in cancer was primarily based on the assumption that increased PKC activation and expression promotes carcinogen induced tumorigenesis [103,150–153].

However, increasing evidence suggests that many PKC isozymes can act as both tumour suppressors and oncogenes. For example, PKC δ has pro-apoptotic effects leading to the belief that it is acting as a tumour suppressor but it has also been linked to the progression of pancreatic and lung cancers [154–156]. Similarly, PKC ζ overexpression in colon cancer cell lines decreases tumour formation in nude mice while loss of PKC ζ is also associated with decreased tumorigenicity [157,158]. Many efforts have been made to create molecules that target PKCs for cancer therapy, but attempts to date have been largely unsuccessful [159]. This may owe to the difficulty in creating inhibitors that target specific PKC isozymes in cancer, coupled with the challenges associated with indirectly disrupting the physiological role of PKCs in normal cells. PKC mutations exist in a diverse range of cancers, existing in the entire coding region with no hotspots. However, it is worth noting that recent pioneering work conducted in the Newton lab surprisingly demonstrated the majority of mutations found in PKCs resulted in a loss of function and none were activating [160]. Further to this, meta-analysis on patients

with non-small cell lung cancer in controlled trials of PKC inhibitors, combined with chemotherapy in comparison with chemotherapy alone, revealed that PKC inhibitors significantly decreased response rates and disease control rates [161].

The ideas presented in this review highlight the challenges in targeting PKCs; clearly defining which PKC isozyme is displaying a tumour suppressor and/or tumour promotor roles and in which tissues proves to be a difficult task. Protein Kinase Cs are key intracellular targets for growth factor and adhesion signaling pathways so perhaps a refocus and revisit of the pathways upstream of PKCs presents a more favourable approach to targeting this group of kinases.

Acknowledgments

This work was supported, in whole by grants received from the Irish Cancer Society, Grant CRS12DOW (to CD) and funds from Science Foundation Ireland, Grant 13/CDA/2228 (to PK). We are grateful to our colleagues in the Laboratory of Cellular and Molecular Biology for helpful discussions and critical review.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Mouw, J.K.; Ou, G.; Weaver, V.M. Extracellular matrix assembly: A multiscale deconstruction. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 771–785. [[CrossRef](#)] [[PubMed](#)]
2. Kim, S.-H.; Turnbull, J.; Guimond, S. Extracellular matrix and cell signalling: The dynamic cooperation of integrin, proteoglycan and growth factor receptor. *J. Endocrinol.* **2011**, *209*, 139–151. [[CrossRef](#)] [[PubMed](#)]
3. Mecham, R.P. Overview of extracellular matrix. In *Current Protocols in Cell Biology*; John Wiley & Sons, Inc.: Hoboken, NJ, USA, 2012; Volume 10.
4. Lu, P.; Weaver, V.M.; Werb, Z. The extracellular matrix: A dynamic niche in cancer progression. *J. Cell Biol.* **2012**, *196*, 395–406. [[CrossRef](#)] [[PubMed](#)]
5. Lemmon, M.A.; Schlessinger, J. Cell signaling by receptor tyrosine kinases. *Cell* **2010**, *141*, 1117–1134. [[CrossRef](#)] [[PubMed](#)]
6. Venkatakrisnan, A.; Deupi, X.; Lebon, G.; Tate, C.G.; Schertler, G.F.; Babu, M.M. Molecular signatures of G-protein-coupled receptors. *Nature* **2013**, *494*, 185–194. [[CrossRef](#)] [[PubMed](#)]
7. Seguin, L.; Desgrosellier, J.S.; Weis, S.M.; Cheresch, D.A. Integrins and cancer: Regulators of cancer stemness, metastasis, and drug resistance. *Trends Cell Biol.* **2015**, *25*, 234–240. [[CrossRef](#)] [[PubMed](#)]
8. Schlessinger, J. Cell signaling by receptor tyrosine kinases. *Cell* **2000**, *103*, 211–225. [[CrossRef](#)]
9. Gschwind, A.; Fischer, O.M.; Ullrich, A. The discovery of receptor tyrosine kinases: Targets for cancer therapy. *Nat. Rev. Cancer* **2004**, *4*, 361–370. [[CrossRef](#)] [[PubMed](#)]
10. Ullrich, A.; Schlessinger, J. Signal transduction by receptors with tyrosine kinase activity. *Cell* **1990**, *61*, 203–212. [[CrossRef](#)]

11. Campbell, I.D.; Humphries, M.J. Integrin structure, activation, and interactions. *Cold Spring Harb Perspect. Biol.* **2011**, *3*. [[CrossRef](#)] [[PubMed](#)]
12. Humphries, J.D.; Byron, A.; Humphries, M.J. Integrin ligands at a glance. *J. Cell Sci.* **2006**, *119*, 3901–3903. [[CrossRef](#)] [[PubMed](#)]
13. Huttenlocher, A.; Horwitz, A.R. Integrins in cell migration. *Cold Spring Harb Perspect. Biol.* **2011**, *3*, a005074. [[CrossRef](#)] [[PubMed](#)]
14. Pawson, T.; Scott, J.D. Signaling through scaffold, anchoring, and adaptor proteins. *Science* **1997**, *278*, 2075–2080. [[CrossRef](#)] [[PubMed](#)]
15. Reiser, J.; Sever, S.; Faul, C. Signal transduction in podocytes—spotlight on receptor tyrosine kinases. *Nat. Rev. Nephrol.* **2014**, *10*, 104–115. [[CrossRef](#)] [[PubMed](#)]
16. Park, J.B.; Lee, C.S.; Jang, J.-H.; Ghim, J.; Kim, Y.-J.; You, S.; Hwang, D.; Suh, P.-G.; Ryu, S.H. Phospholipase signalling networks in cancer. *Nat. Rev. Cancer* **2012**, *12*, 782–792. [[CrossRef](#)] [[PubMed](#)]
17. Marshall, C. Specificity of receptor tyrosine kinase signaling: Transient *versus* sustained extracellular signal-regulated kinase activation. *Cell* **1995**, *80*, 179–185. [[CrossRef](#)]
18. Plopper, G.E.; McNamee, H.P.; Dike, L.E.; Bojanowski, K.; Ingber, D.E. Convergence of integrin and growth factor receptor signaling pathways within the focal adhesion complex. *Mol. Biol. Cell* **1995**, *6*, 1349–1365. [[CrossRef](#)] [[PubMed](#)]
19. Yamada, K.M.; Miyamoto, S. Integrin transmembrane signaling and cytoskeletal control. *Curr. Opin. Cell Biol.* **1995**, *7*, 681–689. [[CrossRef](#)]
20. Zhao, X.; Guan, J.-L. Focal adhesion kinase and its signaling pathways in cell migration and angiogenesis. *Adv. Drug Deliv. Rev.* **2011**, *63*, 610–615. [[CrossRef](#)] [[PubMed](#)]
21. Adams, D.R.; Ron, D.; Kiely, P.A. Rack1, a multifaceted scaffolding protein: Structure and function. *Cell Commun. Signal* **2011**, *9*, 22. [[CrossRef](#)] [[PubMed](#)]
22. Dwane, S.; Durack, E.; O'Connor, R.; Kiely, P.A. Rack1 promotes neurite outgrowth by scaffolding agap2 to fak. *Cell. Signal* **2014**, *26*, 9–18. [[CrossRef](#)] [[PubMed](#)]
23. Kiely, P.A.; Baillie, G.S.; Barrett, R.; Buckley, D.A.; Adams, D.R.; Houslay, M.D.; O'Connor, R. Phosphorylation of rack1 on tyrosine 52 by c-abl is required for insulin-like growth factor i-mediated regulation of focal adhesion kinase. *J. Biol. Chem.* **2009**, *284*, 20263–20274. [[CrossRef](#)] [[PubMed](#)]
24. Kiely, P.A.; Baillie, G.S.; Lynch, M.J.; Houslay, M.D.; O'Connor, R. Tyrosine 302 in rack1 is essential for insulin-like growth factor-i-mediated competitive binding of pp2a and beta1 integrin and for tumor cell proliferation and migration. *J. Biol. Chem.* **2008**, *283*, 22952–22961. [[CrossRef](#)] [[PubMed](#)]
25. Kiely, P.A.; Leahy, M.; O'Gorman, D.; O'Connor, R. Rack1-mediated integration of adhesion and insulin-like growth factor I (IGF-I) signaling and cell migration are defective in cells expressing an IGF-I receptor mutated at tyrosines 1250 and 1251. *J. Biol. Chem.* **2005**, *280*, 7624–7633. [[CrossRef](#)] [[PubMed](#)]
26. Kiely, P.A.; O'Gorman, D.; Luong, K.; Ron, D.; O'Connor, R. Insulin-like growth factor i controls a mutually exclusive association of rack1 with protein phosphatase 2a and β 1 integrin to promote cell migration. *Mol. Cell. Biol.* **2006**, *26*, 4041–4051. [[CrossRef](#)] [[PubMed](#)]

27. Kiely, P.A.; Sant, A.; O'Connor, R. Rack1 is an insulin-like growth factor 1 (IGF-1) receptor-interacting protein that can regulate IGF-1-mediated akt activation and protection from cell death. *J. Biol. Chem.* **2002**, *277*, 22581–22589. [[CrossRef](#)] [[PubMed](#)]
28. O'Donovan, H.C.; Kiely, P.A.; O'Connor, R. Effects of rack1 on cell migration and IGF-I signalling in cardiomyocytes are not dependent on an association with the IGF-IR. *Cell. Signal* **2007**, *19*, 2588–2595. [[CrossRef](#)] [[PubMed](#)]
29. Zeng, L.; Webster, S.; Newton, P. The biology of protein kinase C. In *Calcium Signaling*; Islam, M.S., Ed.; Springer: Dordrecht, The Netherlands, 2012; Volume 740, pp. 639–661.
30. Newton, A.C. Protein kinase C: Structure, function, and regulation. *J. Biol. Chem.* **1995**, *270*, 28495–28498. [[CrossRef](#)] [[PubMed](#)]
31. Newton, A.C. Protein kinase C: Poised to signal. *Am. J. Physiol. Endocrinol. Metab.* **2010**, *298*, E395–E402. [[CrossRef](#)] [[PubMed](#)]
32. Bosco, R.; Melloni, E.; Celeghini, C.; Rimondi, E.; Vaccarezza, M.; Zauli, G. Fine tuning of protein kinase C (PKC) isoforms in cancer: Shortening the distance from the laboratory to the bedside. *Mini Rev. Med. Chem.* **2011**, *11*, 185–199. [[CrossRef](#)] [[PubMed](#)]
33. Mochly-Rosen, D.; Das, K.; Grimes, K.V. Protein kinase C, an elusive therapeutic target? *Nat. Rev. Drug Discov.* **2012**, *11*, 937–957. [[CrossRef](#)] [[PubMed](#)]
34. Ohno, S.; Nishizuka, Y. Protein kinase C isotypes and their specific functions: Prologue. *J. Biochem.* **2002**, *132*, 509–511. [[CrossRef](#)] [[PubMed](#)]
35. Fogh, B.S.; Multhaupt, H.A.B.; Couchman, J.R. Protein kinase C, focal adhesions and the regulation of cell migration. *J. Histochem. Cytochem.* **2014**, *62*, 172–184. [[CrossRef](#)] [[PubMed](#)]
36. Mellor, H.; Parker, P. The extended protein kinase C superfamily. *Biochem. J.* **1998**, *332*, 281–292. [[PubMed](#)]
37. Martiny-Baron, G.; Fabbro, D. Classical PKC isoforms in cancer. *Pharmacol. Res.* **2007**, *55*, 477–486. [[CrossRef](#)] [[PubMed](#)]
38. Poli, A.; Mongiorgi, S.; Cocco, L.; Follo, M.Y. Protein kinase C involvement in cell cycle modulation. *Biochem. Soc. Trans.* **2014**, *42*, 1471–1476. [[CrossRef](#)] [[PubMed](#)]
39. Defilippi, P.; Venturino, M.; Gulino, D.; Duperray, A.; Boquet, P.; Fiorentini, C.; Volpe, G.; Palmieri, M.; Silengo, L.; Tarone, G. Dissection of pathways implicated in integrin-mediated actin cytoskeleton assembly involvement of protein kinase C, RHO GTPASE, and tyrosine phosphorylation. *J. Biol. Chem.* **1997**, *272*, 21726–21734. [[CrossRef](#)] [[PubMed](#)]
40. Huang, X.; Wu, J.; Spong, S.; Sheppard, D. The integrin α v β 6 is critical for keratinocyte migration on both its known ligand, fibronectin, and on vitronectin. *J. Cell Sci.* **1998**, *111*, 2189–2195. [[PubMed](#)]
41. Bordeleau, F.; Galarneau, L.; Gilbert, S.; Loranger, A.; Marceau, N. Keratin 8/18 modulation of protein kinase C-mediated integrin-dependent adhesion and migration of liver epithelial cells. *Mol. Biol. Cell* **2010**, *21*, 1698–1713. [[CrossRef](#)] [[PubMed](#)]
42. Gonelli, A.; Mischiati, C.; Guerrini, R.; Voltan, R.; Salvadori, S.; Zauli, G. Perspectives of protein kinase C (PKC) inhibitors as anti-cancer agents. *Mini Rev. Med. Chem.* **2009**, *9*, 498–509. [[CrossRef](#)] [[PubMed](#)]

43. Zhang, W.; Zong, C.S.; Hermanto, U.; Lopez-Bergami, P.; Ronai, Z.E.; Wang, L.-H. Rack1 recruits STAT3 specifically to insulin and insulin-like growth factor 1 receptors for activation, which is important for regulating anchorage-independent growth. *Mol. Cell. Biol.* **2006**, *26*, 413–424. [[CrossRef](#)] [[PubMed](#)]
44. Haimovich, B.; Kaneshiki, N.; Ji, P. Protein kinase C regulates tyrosine phosphorylation of PP125FAK in platelets adherent to fibrinogen. *Blood* **1996**, *87*, 152–161. [[PubMed](#)]
45. Steinberg, S.F. Structural basis of protein kinase C isoform function. *Physiol. Rev.* **2008**, *88*, 1341–1378. [[CrossRef](#)] [[PubMed](#)]
46. Rosse, C.; Linch, M.; Kermorgant, S.; Cameron, A.J.M.; Boeckeler, K.; Parker, P.J. PKC and the control of localized signal dynamics. *Nat. Rev. Mol. Cell. Biol.* **2010**, *11*, 103–112. [[CrossRef](#)] [[PubMed](#)]
47. Antal, C.E.; Violin, J.D.; Kunkel, M.T.; SkovsÅy, S.; Newton, A.C. Intramolecular conformational changes optimize protein kinase C signaling. *Chem. Biol.* **2014**, *21*, 459–469. [[CrossRef](#)] [[PubMed](#)]
48. ColÅšn-GonzÅąlez, F.; Kazanietz, M.G. C1 domains exposed: From diacylglycerol binding to protein–protein interactions. *Biochim. Biophys. Acta* **2006**, *1761*, 827–837. [[CrossRef](#)] [[PubMed](#)]
49. Cho, W. Membrane targeting by C1 and C2 domains. *J. Biol. Chem.* **2001**, *276*, 32407–32410. [[CrossRef](#)] [[PubMed](#)]
50. Ananthanarayanan, B.; Stahelin, R.V.; Digman, M.A.; Cho, W. Activation mechanisms of conventional protein kinase C isoforms are determined by the ligand affinity and conformational flexibility of their C1 domains. *J. Biol. Chem.* **2003**, *278*, 46886–46894. [[CrossRef](#)] [[PubMed](#)]
51. Stahelin, R.V. Ready, set, go! How protein kinase c manages dynamic signaling. *Chem. Biol.* **2014**, *21*, 433–434. [[CrossRef](#)] [[PubMed](#)]
52. Oancea, E.; Meyer, T. Protein kinase C as a molecular machine for decoding calcium and diacylglycerol signals. *Cell* **1998**, *95*, 307–318. [[CrossRef](#)]
53. Newton, A. Regulation of the abc kinases by phosphorylation: Protein kinase C as a paradigm. *Biochem. J.* **2003**, *370*, 361–371. [[CrossRef](#)] [[PubMed](#)]
54. Messerschmidt, A.; Macieira, S.; Velarde, M.; BÅd’deker, M.; Benda, C.; Jestel, A.; Brandstetter, H.; Neufelind, T.; Blaessee, M. Crystal structure of the catalytic domain of human atypical protein kinase C-IOTA reveals interaction mode of phosphorylation site in turn motif. *J. Mol. Biol.* **2005**, *352*, 918–931. [[CrossRef](#)] [[PubMed](#)]
55. Seki, T.; Matsubayashi, H.; Amano, T.; Shirai, Y.; Saito, N.; Sakai, N. Phosphorylation of PKC activation loop plays an important role in receptor-mediated translocation of PKC. *Genes Cells* **2005**, *10*, 225–239. [[CrossRef](#)] [[PubMed](#)]
56. Dutil, E.M.; Newton, A.C. Dual role of pseudosubstrate in the coordinated regulation of protein kinase C by phosphorylation and diacylglycerol. *J. Biol. Chem.* **2000**, *275*, 10697–10701. [[CrossRef](#)] [[PubMed](#)]
57. Hauge, C.; Antal, T.L.; Hirschberg, D.; Doehn, U.; Thorup, K.; Idrissova, L.; Hansen, K.; Jensen, O.N.; JÅyrgensen, T.J.; Biondi, R.M. Mechanism for activation of the growth factor-activated AGC kinases by turn motif phosphorylation. *EMBO J.* **2007**, *26*, 2251–2261. [[CrossRef](#)] [[PubMed](#)]

58. Behn-Krappa, A.; Newton, A.C. The hydrophobic phosphorylation motif of conventional protein kinase C is regulated by autophosphorylation. *Curr. Biol.* **1999**, *9*, 728–737. [[CrossRef](#)]
59. Oliva, J.L.; Griner, E.M.; Kazanietz, M.G. PKC isozymes and diacylglycerol-regulated proteins as effectors of growth factor receptors. *Growth Factors* **2005**, *23*, 245–252. [[CrossRef](#)] [[PubMed](#)]
60. Smith, M.R.; DeGudicibus, S.J.; Stacey, D.W. Requirement for C-RAS proteins during viral oncogene transformation. *Nature* **1986**, *320*, 540–543. [[CrossRef](#)] [[PubMed](#)]
61. Margolis, B.; Rhee, S.; Felder, S.; Mervic, M.; Lyall, R.; Levitzki, A.; Ullrich, A.; Zilberstein, A.; Schlessinger, J. EGF induces tyrosine phosphorylation of phospholipase C-II: A potential mechanism for EGF receptor signaling. *Cell* **1989**, *57*, 1101–1107. [[CrossRef](#)]
62. Lemmon, M.A.; Schlessinger, J. Regulation of signal transduction and signal diversity by receptor oligomerization. *Trends Biochem. Sci.* **1994**, *19*, 459–463. [[CrossRef](#)]
63. Vivanco, I.; Sawyers, C.L. The phosphatidylinositol 3-kinase–AKT pathway in human cancer. *Nat. Rev. Cancer* **2002**, *2*, 489–501. [[CrossRef](#)] [[PubMed](#)]
64. Domin, J.; Waterfield, M.D. Using structure to define the function of phosphoinositide 3-kinase family members. *FEBS Lett.* **1997**, *410*, 91–95. [[CrossRef](#)]
65. White, M.F. The IRS-signalling system: A network of docking proteins that mediate insulin action. *Mol. Cell. Biochem.* **1998**, *182*, 3–11. [[CrossRef](#)] [[PubMed](#)]
66. Vanhaesebroeck, B.; Stephens, L.; Hawkins, P. PI3K signalling: The path to discovery and understanding. *Nat. Rev. Mol. Cell Biol.* **2012**, *13*, 195–203. [[CrossRef](#)] [[PubMed](#)]
67. Williams, R.L.; Katan, M. Structural views of phosphoinositide-specific phospholipase C: Signalling the way ahead. *Structure* **1996**, *4*, 1387–1394. [[CrossRef](#)]
68. Todderud, G.; Wahl, M.I.; Rhee, S.G.; Carpenter, G. Stimulation of phospholipase C- γ 1 membrane association by epidermal growth factor. *Science* **1990**, *249*, 296–298. [[CrossRef](#)] [[PubMed](#)]
69. Nishizuka, Y. Intracellular signaling by hydrolysis of phospholipids and activation of protein kinase C. *Science* **1992**, *258*, 607–614. [[CrossRef](#)] [[PubMed](#)]
70. Peters, K.G.; Marie, J.; Wilson, E.; Ives, H.E.; Escobedo, J.; Rosario, M.D.; Mirda, D.; Williams, L.T. Point mutation of an FGF receptor abolishes phosphatidylinositol turnover and Ca^{2+} flux but not mitogenesis. *Nature* **1992**, *358*, 678–681. [[CrossRef](#)] [[PubMed](#)]
71. Nishibe, S.; Wahl, M.I.; Wedegaertner, P.B.; Kim, J.; Rhee, S.G.; Carpenter, G.; Kim, J. Selectivity of phospholipase C phosphorylation by the epidermal growth factor receptor, the insulin receptor, and their cytoplasmic domains. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 424–428. [[CrossRef](#)] [[PubMed](#)]
72. Moriya, S.; Kazlauskas, A.; Akimoto, K.; Hirai, S.; Mizuno, K.; Takenawa, T.; Fukui, Y.; Watanabe, Y.; Ozaki, S.; Ohno, S. Platelet-derived growth factor activates protein kinase C ϵ through redundant and independent signaling pathways involving phospholipase C γ or phosphatidylinositol 3-kinase. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 151–155. [[CrossRef](#)] [[PubMed](#)]
73. Larose, L.; Gish, G.; Shoelson, S.; Pawson, T. Identification of residues in the beta platelet-derived growth factor receptor that confer specificity for binding to phospholipase C- γ 1. *Oncogene* **1993**, *8*, 2493–2499. [[PubMed](#)]

74. Akimoto, K.; Takahashi, R.; Moriya, S.; Nishioka, N.; Takayanagi, J.; Kimura, K.; Fukui, Y.; Osada, S.I.; Mizuno, K.; Hirai, S.I.; *et al.* EGF or PDGF receptors activate atypical pklambda through phosphatidylinositol 3-kinase. *EMBO J.* **1996**, *15*, 788–798. [[PubMed](#)]
75. Brodie, C.; Bogi, K.; Acs, P.; Lazarovici, P.; Petrovics, G.; Anderson, W.B.; Blumberg, P.M. Protein kinase C-epsilon plays a role in neurite outgrowth in response to epidermal growth factor and nerve growth factor in PC12 cells. *Cell Growth Differ.* **1999**, *10*, 183–191. [[PubMed](#)]
76. Greco, S.; Muscella, A.; Elia, M.G.; Salvatore, P.; Storelli, C.; Mazzotta, A.; Manca, C.; Marsigliante, S. Angiotensin II activates extracellular signal regulated kinases via protein kinase C and epidermal growth factor receptor in breast cancer cells. *J. Cell. Physiol.* **2003**, *196*, 370–377. [[CrossRef](#)] [[PubMed](#)]
77. Burry, R.W. PKC activators (phorbol ester or bryostatin) stimulate outgrowth of ngf-dependent neurites in a subline of PC12 cells. *J. Neurosc. Res.* **1998**, *53*, 214–222. [[CrossRef](#)]
78. Welsh, J.B.; Gill, G.N.; Rosenfeld, M.G.; Wells, A. A negative feedback loop attenuates EGF-induced morphological changes. *J. Cell Biol.* **1991**, *114*, 533–543. [[CrossRef](#)] [[PubMed](#)]
79. Morrison, P.; Takishima, K.; Rosner, M.R. Role of threonine residues in regulation of the epidermal growth factor receptor by protein kinase C and mitogen-activated protein kinase. *J. Biol. Chem.* **1993**, *268*, 15536–15543. [[PubMed](#)]
80. Lund, K.; Lazar, C.S.; Chen, W.; Walsh, B.; Welsh, J.; Herbst, J.; Walton, G.; Rosenfeld, M.; Gill, G.; Wiley, H. Phosphorylation of the epidermal growth factor receptor at threonine 654 inhibits ligand-induced internalization and down-regulation. *J. Biol. Chem.* **1990**, *265*, 20517–20523. [[PubMed](#)]
81. Cochet, C.; Gill, G.N.; Meisenhelder, J.; Cooper, J.A.; Hunter, T. C-kinase phosphorylates the epidermal growth factor receptor and reduces its epidermal growth factor-stimulated tyrosine protein kinase activity. *J. Biol. Chem.* **1984**, *259*, 2553–2558. [[PubMed](#)]
82. Chen, P.; Xie, H.; Wells, A. Mitogenic signaling from the EGF receptor is attenuated by a phospholipase C- γ /protein kinase C feedback mechanism. *Mol. Biol. Cell.* **1996**, *7*, 871–881. [[CrossRef](#)] [[PubMed](#)]
83. Hackel, P.O.; Zwick, E.; Prenzel, N.; Ullrich, A. Epidermal growth factor receptors: Critical mediators of multiple receptor pathways. *Curr. Opin. Cell Biol.* **1999**, *11*, 184–189. [[CrossRef](#)]
84. Chen, P.; Xie, H.; Sekar, M.C.; Gupta, K.; Wells, A. Epidermal growth factor receptor-mediated cell motility: Phospholipase C activity is required, but mitogen-activated protein kinase activity is not sufficient for induced cell movement. *J. Cell Biol.* **1994**, *127*, 847–857. [[CrossRef](#)] [[PubMed](#)]
85. Kermorgant, S.; Zicha, D.; Parker, P.J. Protein kinase C controls microtubule-based traffic but not proteasomal degradation of C-MET. *J. Biol. Chem.* **2003**, *278*, 28921–28929. [[CrossRef](#)] [[PubMed](#)]
86. Kermorgant, S.; Zicha, D.; Parker, P.J. PKC controls HGF-dependent C-MET traffic, signalling and cell migration. *EMBO J.* **2004**, *23*, 3721–3734. [[CrossRef](#)] [[PubMed](#)]
87. Besson, A.; Davy, A.; Robbins, S.M.; Yong, V.W. Differential activation of ERKS to focal adhesions by PKC epsilon is required for PMA-induced adhesion and migration of human glioma cells. *Oncogene* **2001**, *20*, 7398–7407. [[CrossRef](#)] [[PubMed](#)]

88. Hashigasako, A.; Machide, M.; Nakamura, T.; Matsumoto, K.; Nakamura, T. Bi-directional regulation of SER-985 phosphorylation of C-met via protein kinase c and protein phosphatase 2a involves C-met activation and cellular responsiveness to hepatocyte growth factor. *J. Biol. Chem.* **2004**, *279*, 26445–26452. [[CrossRef](#)] [[PubMed](#)]
89. Dorsam, R.T.; Gutkind, J.S. G-protein-coupled receptors and cancer. *Nat. Rev. Cancer* **2007**, *7*, 79–94. [[CrossRef](#)] [[PubMed](#)]
90. Gilman, A.G. G proteins: Transducers of receptor-generated signals. *Annu. Rev. Biochem.* **1987**, *56*, 615–649. [[CrossRef](#)] [[PubMed](#)]
91. O’Hayre, M.; Vazquez-Prado, J.; Kufareva, I.; Stawiski, E.W.; Handel, T.M.; Seshagiri, S.; Gutkind, J.S. The emerging mutational landscape of G proteins and G-protein-coupled receptors in cancer. *Nat. Rev. Cancer* **2013**, *13*, 412–424. [[CrossRef](#)] [[PubMed](#)]
92. Neves, S.R.; Ram, P.T.; Iyengar, R. G protein pathways. *Science* **2002**, *296*, 1636–1639. [[CrossRef](#)] [[PubMed](#)]
93. Milligan, G.; Kostenis, E. Heterotrimeric G-proteins: A short history. *Br. J. pharmacol.* **2006**, *147*, S46–S55. [[CrossRef](#)] [[PubMed](#)]
94. Dupr  l, D.J.; Robitaille, M.; Rebois, R.V.; H  lbert, T.E. The role of G    subunits in the organization, assembly, and function of gpcr signaling complexes. *Annu. Rev. pharmacol. Toxicol.* **2009**, *49*, 31–56. [[CrossRef](#)] [[PubMed](#)]
95. Kolanus, W.; Seed, B. Integrins and inside-out signal transduction: Converging signals from PKC and PIP 3. *Curr. Opin. Cell Biol.* **1997**, *9*, 725–731. [[CrossRef](#)]
96. Parekh, D.B.; Ziegler, W.; Parker, P.J. Multiple pathways control protein kinase C phosphorylation. *EMBO J.* **2000**, *19*, 496–503. [[CrossRef](#)] [[PubMed](#)]
97. Disatnik, M.H.; Hernandez-Sotomayor, S.M.; Jones, G.; Carpenter, G.; Mochly-Rosen, D. Phospholipase C-gamma 1 binding to intracellular receptors for activated protein kinase C. *Proc. Natl. Acad. Sci. USA* **1994**, *91*, 559–563. [[CrossRef](#)] [[PubMed](#)]
98. Ginsberg, M.H.; Du, X.; Plow, E.F. Inside-out integrin signalling. *Curr. Opin. Cell Biol.* **1992**, *4*, 766–771. [[CrossRef](#)]
99. Ivashkiv, L.B. Cross-regulation of signaling by itam-associated receptors. *Nat. Immunol.* **2009**, *10*, 340–347. [[CrossRef](#)] [[PubMed](#)]
100. Woods, A.; Couchman, J.R. Protein kinase C involvement in focal adhesion formation. *J. Cell Sci.* **1992**, *101*, 277–290. [[PubMed](#)]
101. Griner, E.M.; Kazanietz, M.G. Protein kinase C and other diacylglycerol effectors in cancer. *Nat. Rev. Cancer* **2007**, *7*, 281–294. [[CrossRef](#)] [[PubMed](#)]
102. Ron, D.; Kazanietz, M.G. New insights into the regulation of protein kinase C and novel phorbol ester receptors. *FASEB J.* **1999**, *13*, 1658–1676. [[PubMed](#)]
103. Barr, L.F.; Mabry, M.; Nelkin, B.D.; Tyler, G.; May, W.S.; Baylin, S.B. C-MYC gene-induced alterations in protein kinase C expression: A possible mechanism facilitating MYC-RAS gene complementation. *Cancer Res.* **1991**, *51*, 5514–5519. [[PubMed](#)]
104. Han, E.K.-H.; Cacace, A.M.; Sgambato, A.; Weinstein, I.B. Altered expression of cyclins and C-FOS in R6 cells that overproduce PKC  . *Carcinogenesis* **1995**, *16*, 2423–2428. [[CrossRef](#)] [[PubMed](#)]

105. Da Rocha, A.B.; Mans, D.R.A.; Regner, A.; Schwartzmann, G. Targeting protein kinase C: New therapeutic opportunities against high-grade malignant gliomas? *Oncologist* **2002**, *7*, 17–33. [[CrossRef](#)] [[PubMed](#)]
106. Lu, Z.; Hornia, A.; Jiang, Y.W.; Zang, Q.; Ohno, S.; Foster, D.A. Tumor promotion by depleting cells of protein kinase C delta. *Mol. Cell. Biol.* **1997**, *17*, 3418–3428. [[PubMed](#)]
107. Lu, Z.; Liu, D.; Hornia, A.; Devonish, W.; Pagano, M.; Foster, D.A. Activation of protein kinase C triggers its ubiquitination and degradation. *Mol. Cell. Biol.* **1998**, *18*, 839–845. [[PubMed](#)]
108. Gould, C.M.; Newton, A.C. The life and death of protein kinase c. *Curr. Drug Targets* **2008**, *9*, 614–625. [[CrossRef](#)] [[PubMed](#)]
109. Koivunen, J.; Aaltonen, V.; Peltonen, J. Protein kinase C (PKC) family in cancer progression. *Cancer Lett.* **2006**, *235*, 1–10. [[CrossRef](#)] [[PubMed](#)]
110. Langzam, L.; Koren, R.; Gal, R.; Kugel, V.; Paz, A.; Farkas, A.; Sampson, S.R. Patterns of protein kinase C isoenzyme expression in transitional cell carcinoma of bladder. Relation to degree of malignancy. *Am. J. Clin. Pathol.* **2001**, *116*, 377–385. [[CrossRef](#)] [[PubMed](#)]
111. Varga, A.; Czifra, G.; Tallai, B.; Nemeth, T.; Kovacs, I.; Kovacs, L.; Biro, T. Tumor grade-dependent alterations in the protein kinase C isoform pattern in urinary bladder carcinomas. *Eur. Urol.* **2004**, *46*, 462–465. [[CrossRef](#)] [[PubMed](#)]
112. Koren, R.; Ben Meir, D.; Langzam, L.; Dekel, Y.; Konichezky, M.; Baniel, J.; Livne, P.M.; Gal, R.; Sampson, S.R. Expression of protein kinase C isoenzymes in benign hyperplasia and carcinoma of prostate. *Oncol. Rep.* **2004**, *11*, 321–326. [[CrossRef](#)] [[PubMed](#)]
113. Fournier, D.B.; Chisamore, M.; Lurain, J.R.; Rademaker, A.W.; Jordan, V.C.; Tonetti, D.A. Protein kinase C α expression is inversely related to er status in endometrial carcinoma: Possible role in AP-1-mediated proliferation of ER-negative endometrial cancer. *Gynecol. Oncol.* **2001**, *81*, 366–372. [[CrossRef](#)] [[PubMed](#)]
114. Tsai, J.H.; Tsai, M.T.; Su, W.W.; Chen, Y.L.; Wu, T.T.; Hsieh, Y.S.; Huang, C.Y.; Yeh, K.T.; Liu, J.Y. Expression of protein kinase C alpha in biopsies and surgical specimens of human hepatocellular carcinoma. *Chin. J. Physiol.* **2005**, *48*, 139–143. [[PubMed](#)]
115. Neill, G.W.; Ghali, L.R.; Green, J.L.; Ikram, M.S.; Philpott, M.P.; Quinn, A.G. Loss of protein kinase calpha expression may enhance the tumorigenic potential of GLI1 in basal cell carcinoma. *Cancer Res.* **2003**, *63*, 4692–4697. [[PubMed](#)]
116. Kahl-Rainer, P.; Karner-Hanusch, J.; Weiss, W.; Marian, B. Five of six protein kinase C isoenzymes present in normal mucosa show reduced protein levels during tumor development in the human colon. *Carcinogenesis* **1994**, *15*, 779–782. [[CrossRef](#)] [[PubMed](#)]
117. Kerfoot, C.; Huang, W.; Rotenberg, S.A. Immunohistochemical analysis of advanced human breast carcinomas reveals downregulation of protein kinase C α . *J. Histochem. Cytochem.* **2004**, *52*, 419–422. [[CrossRef](#)] [[PubMed](#)]
118. Ways, D.K.; Kukoly, C.A.; deVente, J.; Hooker, J.L.; Bryant, W.O.; Posekany, K.J.; Fletcher, D.J.; Cook, P.P.; Parker, P.J. MCF-7 breast cancer cells transfected with protein kinase C- α exhibit altered expression of other protein kinase C isoforms and display a more aggressive neoplastic phenotype. *J. Clin. Investig.* **1995**, *95*, 1906–1915. [[CrossRef](#)] [[PubMed](#)]

132. Lee, K.-W.; Kim, S.G.; Kim, H.-P.; Kwon, E.; You, J.; Choi, H.-J.; Park, J.-H.; Kang, B.-C.; Im, S.-A.; Kim, T.-Y. Enzastaurin, a protein kinase C β inhibitor, suppresses signaling through the ribosomal S6 kinase and bad pathways and induces apoptosis in human gastric cancer cells. *Cancer Res.* **2008**, *68*, 1916–1926. [[CrossRef](#)] [[PubMed](#)]
133. Yoshiji, H.; Kuriyama, S.; Ways, D.K.; Yoshii, J.; Miyamoto, Y.; Kawata, M.; Ikenaka, Y.; Tsujinoue, H.; Nakatani, T.; Shibuya, M. Protein kinase c lies on the signaling pathway for vascular endothelial growth factor-mediated tumor development and angiogenesis. *Cancer Res.* **1999**, *59*, 4413–4418. [[PubMed](#)]
134. Xia, P.; Aiello, L.P.; Ishii, H.; Jiang, Z.Y.; Park, D.J.; Robinson, G.S.; Takagi, H.; Newsome, W.P.; Jirousek, M.R.; King, G.L. Characterization of vascular endothelial growth factor's effect on the activation of protein kinase C, its isoforms, and endothelial cell growth. *J. Clin. Investig.* **1996**, *98*, 2018–2026. [[CrossRef](#)] [[PubMed](#)]
135. Kamimura, K.; Hojo, H.; Abe, M. Characterization of expression of protein kinase c isozymes in human b-cell lymphoma: Relationship between its expression and prognosis. *Pathol. Int.* **2004**, *54*, 224–230. [[CrossRef](#)] [[PubMed](#)]
136. Mazzoni, E.; Adam, A.; de Kier Joffe, E.B.; Aguirre-Ghiso, J.A. Immortalized mammary epithelial cells overexpressing protein kinase C γ acquire a malignant phenotype and become tumorigenic *in vivo*. *Mol. Cancer Res.* **2003**, *1*, 776–787. [[PubMed](#)]
137. Swannie, H.; Kaye, S. Protein kinase c inhibitors. *Curr. Oncol. Rep.* **2002**, *4*, 37–46. [[CrossRef](#)] [[PubMed](#)]
138. Gray, M.O.; Karliner, J.S.; Mochly-Rosen, D. A selective ε -protein kinase C antagonist inhibits protection of cardiac myocytes from hypoxia-induced cell death. *J. Biol. Chem.* **1997**, *272*, 30945–30951. [[CrossRef](#)] [[PubMed](#)]
139. Leskow, F.C.; Krasnapolski, M.A.; Urtreger, A.J. The pros and cons of targeting protein kinase C (PKC) in the management of cancer patients. *Curr. Pharm. Biotechnol.* **2011**, *12*, 1961–1973. [[CrossRef](#)] [[PubMed](#)]
140. Andrejauskas-Buchdunger, E.; Regenass, U. Differential inhibition of the epidermal growth factor-, platelet-derived growth factor-, and protein kinase C-mediated signal transduction pathways by the staurosporine derivative CGP 41251. *Cancer Res.* **1992**, *52*, 5353–5358. [[PubMed](#)]
141. Tenzer, A.; Zingg, D.; Rocha, S.; Hemmings, B.; Fabbro, D.; Glanzmann, C.; Schubiger, P.A.; Bodis, S.; Pruschy, M. The phosphatidylinositide 3 \hat{E} -kinase/akt survival pathway is a target for the anticancer and radiosensitizing agent PKC412, an inhibitor of protein kinase c. *Cancer Res.* **2001**, *61*, 8203–8210. [[PubMed](#)]
142. Faul, M.M.; Gillig, J.R.; Jirousek, M.R.; Ballas, L.M.; Schotten, T.; Kahl, A.; Mohr, M. Acyclic *n*-(azacycloalkyl) bisindolylmaleimides: Isozyme selective inhibitors of PKC β . *Bioorg. Med. Chem. Lett.* **2003**, *13*, 1857–1859. [[CrossRef](#)]
143. Mendel, D.B.; Laird, A.D.; Xin, X.; Louie, S.G.; Christensen, J.G.; Li, G.; Schreck, R.E.; Abrams, T.J.; Ngai, T.J.; Lee, L.B.; *et al.* In vivo antitumor activity of su11248, a novel tyrosine kinase inhibitor targeting vascular endothelial growth factor and platelet-derived growth factor

- receptors: Determination of a pharmacokinetic/pharmacodynamic relationship. *Clin. Cancer Res.* **2003**, *9*, 327–337. [[PubMed](#)]
144. Kreisl, T.N.; Kotliarova, S.; Butman, J.A.; Albert, P.S.; Kim, L.; Musib, L.; Thornton, D.; Fine, H.A. A phase I/II trial of enzastaurin in patients with recurrent high-grade gliomas. *Neuro-oncology* **2010**, *12*, 181–189. [[CrossRef](#)] [[PubMed](#)]
145. Workman, P.; Kaye, S.B.; Schwartzmann, G. Laboratory and phase I studies of new cancer drugs. *Curr. Opin. Oncol.* **1992**, *4*, 1065–1072. [[CrossRef](#)] [[PubMed](#)]
146. Pavlick, A.; Wu, J.; Roberts, J.; Rosenthal, M.; Hamilton, A.; Wadler, S.; Farrell, K.; Carr, M.; Fry, D.; Murgo, A.; *et al.* Phase I study of bryostatin 1, a protein kinase C modulator, preceding cisplatin in patients with refractory non-hematologic tumors. *Cancer Chemother. Pharmacol.* **2009**, *64*, 803–810. [[CrossRef](#)] [[PubMed](#)]
147. Souroujon, M.C.; Mochly-Rosen, D. Peptide modulators of protein–protein interactions in intracellular signaling. *Nat. Biotechnol.* **1998**, *16*, 919–924. [[CrossRef](#)] [[PubMed](#)]
148. Kim, J.; Choi, Y.-L.; Vallentin, A.; Hunrichs, B.S.; Hellerstein, M.K.; Peehl, D.M.; Mochly-Rosen, D. Centrosomal PKC β II and pericentrin are critical for human prostate cancer growth and angiogenesis. *Cancer Res.* **2008**, *68*, 6831–6839. [[CrossRef](#)] [[PubMed](#)]
149. Ferreira, J.C.; Koyanagi, T.; Palaniyandi, S.S.; Fajardo, G.; Churchill, E.N.; Budas, G.; Disatnik, M.-H.; Bernstein, D.; Brum, P.C.; Mochly-Rosen, D. Pharmacological inhibition of β IIIPKC is cardioprotective in late-stage hypertrophy. *J. Mol. Cell. Cardiol.* **2011**, *51*, 980–987. [[CrossRef](#)] [[PubMed](#)]
150. Castagna, M.; Takai, Y.; Kaibuchi, K.; Sano, K.; Kikkawa, U.; Nishizuka, Y. Direct activation of calcium-activated, phospholipid-dependent protein kinase by tumor-promoting phorbol esters. *J. Biol. Chem.* **1982**, *257*, 7847–7851. [[PubMed](#)]
151. O’Brian, C.A.; Liskamp, R.M.; Solomon, D.H.; Weinstein, I.B. Inhibition of protein kinase C by tamoxifen. *Cancer Res.* **1985**, *45*, 2462–2465. [[PubMed](#)]
152. Kikkawa, U.; Takai, Y.; Tanaka, Y.; Miyake, R.; Nishizuka, Y. Protein kinase C as a possible receptor protein of tumor-promoting phorbol esters. *J. Biol. Chem.* **1983**, *258*, 11442–11445. [[PubMed](#)]
153. Blobe, G.C.; Obeid, L.M.; Hannun, Y.A. Regulation of protein kinase C and role in cancer biology. *Cancer Metastasis Rev.* **1994**, *13*, 411–431. [[CrossRef](#)] [[PubMed](#)]
154. Reyland, M. Protein kinase cdelta and apoptosis. *Biochem. Soc. Trans.* **2007**, *35*, 1001–04. [[PubMed](#)]
155. Symonds, J.M.; Ohm, A.M.; Carter, C.J.; Heasley, L.E.; Boyle, T.A.; Franklin, W.A.; Reyland, M.E. Protein kinase C δ is a downstream effector of oncogenic k-ras in lung tumors. *Cancer Res.* **2011**, *71*, 2087–2097. [[CrossRef](#)] [[PubMed](#)]
156. Mauro, L.V.; Grossoni, V.C.; Urtreger, A.J.; Yang, C.; Colombo, L.L.; Morandi, A.; Pallotta, M.G.; Kazanietz, M.G.; de Kier Joffe, E.D.B.; Puricelli, L.L. PKC delta (PKC δ) promotes tumoral progression of human ductal pancreatic cancer. *Pancreas* **2010**, *39*, e31–e41. [[CrossRef](#)] [[PubMed](#)]

157. Luna-Ulloa, L.B.; Hernandez-Maqueda, J.G.; Santoyo-Ramos, P.; Castaneda-Patlan, M.C.; Robles-Flores, M. Protein kinase C ζ is a positive modulator of canonical wnt signaling pathway in tumoral colon cell lines. *Carcinogenesis* **2011**, *32*, 1615–1624. [[CrossRef](#)] [[PubMed](#)]
158. Ma, L.; Tao, Y.; Duran, A.; Llado, V.; Galvez, A.; Barger, J.F.; Castilla, E.A.; Chen, J.; Yajima, T.; Porollo, A. Control of nutrient stress-induced metabolic reprogramming by PKC ζ in tumorigenesis. *Cell* **2013**, *152*, 599–611. [[CrossRef](#)] [[PubMed](#)]
159. Mackay, H.J.; Twelves, C.J. Targeting the protein kinase c family: Are we there yet? *Nat. Rev. Cancer* **2007**, *7*, 554–562. [[CrossRef](#)] [[PubMed](#)]
160. Antal, C.E.; Hudson, A.M.; Kang, E.; Zanca, C.; Wirth, C.; Stephenson, N.L.; Trotter, E.W.; Gallegos, L.L.; Miller, C.J.; Furnari, F.B. Cancer-associated protein kinase c mutations reveal kinase's role as tumor suppressor. *Cell* **2015**, *160*, 489–502. [[CrossRef](#)] [[PubMed](#)]
161. Zhang, L.; Cao, F.; Wang, Y.; Meng, F.; Zhang, Y.; Zhong, D.; Zhou, Q. The Protein Kinase C (PKC) inhibitors combined with chemotherapy in the treatment of advanced non-small cell lung cancer: Meta-analysis of randomized controlled trials. *Clin. Trans. Oncol.* **2015**, *17*, 371–377. [[CrossRef](#)] [[PubMed](#)]